

Biomarker Development: How Mass Spectrometry Is Changing the Field

Regulated Bioanalysis Interest Group (RBIG)

5:45-7:00 pm, Wednesday, 07-June-2023

Presiding:

Wenkui Li (Novartis Institutes for BioMedical Research), Jian Wang (Crinetics Pharmaceuticals)

Panelists:

Long Geng, MS (Frontage Laboratories), Aiying Yu, Ph.D. (Genentech), Yifan Shi, Ph.D. (Janssen Research & Development), Yang (Thomas) Tang, Ph.D. (Genentech)



Agenda



- *05:45 05:50 pm Introduction*
- 05:50 06:00 pm Using LC-MS for biomarker analysis: When? How? Why?
- Long Geng, MS (Frontage Laboratories)
- 06:00 06:10 pm Target occupancy in frozen and FFPE tissues using immunoaffinity capture 2D-LC-MS/MS
- Aiying Yu, Ph. D. (Genentech)
- 06:10 06:20 pm -Implementing antibody-free strategies for protein quantitation in monkey liver tissues using liquid chromatography coupled with mass spectrometry
- Yifan Shi, Ph. D. (Janssen Research & Development)
- 06:20 06:30 pm A universal surrogate matrix assay for urea measurement in clinical pharmacokinetic studies of respiratory diseases
- Yang (Thomas) Tang, Ph.D. (Genentech)
- 06:30 7:00 pm Panel discussion







Long Geng, M.S., Ph.D. Candidate

Lab Manager in Frontage Lab Biomarker Department since 2022. M.S. in Bioengineering/ Biomedical Engineering from New Jersey Institute of Technology. Ph.D. Candidate in Biochemical Engineering from Villanova University. Before joining Frontage Lab, he worked at Janssen Research and Development and Teva pharmaceuticals. 10 years of experience in both regulated and non-regulated PK/PD/Biomarker/ADA assay development using ligand binding, LC-MS/MS and hybrid platforms.



USING LC-MS FOR BIOMARKER ANALYSIS: WHEN? HOW? WHY?

Long Geng, Zhongping John Lin Frontage Laboratories

71th ASMS Conference, RBIG Workshop June 7th, 2023



Using LC-MS for biomarker analysis

When? How? Why?

- Bioanalysis assay platform considerations
 - Assay capability between phases
 - Assay challenges and complexity
 - Time and associated costs
- Biomarker assay platforms
 - Immunoassays (MSD, Simoa, ELLA, SMC, Olink)
 - Mass Spectrometry (LC-MS, GC-MS)
 - Genomics (PCR, NGS)
 - Imaging (X-ray, MRI)
 - Flow Cytometry



Using LC-MS for biomarker analysis When? How? Why?



- High assay sensitivity
 - pg/mL to ng/mL level
- Less reagent variability
 - Lot to lot variation
- Less Matrix interference
 - Selectivity, Specificity
 - Different matrices
- Multiplexing
 - 10+ analytes can be analyzed simultaneously
 - Therapeutic target and ligands

Using LC-MS for biomarker analysis

When? How? Why?

Improvement of assay selectivity, specificity, and sensitivity

- Case study 1: Development and validation of an LC–MS/MS Method for the quantitation of heparan sulfate in human urine¹
- Case study 2: A novel LC–MS/MS assay to quantify dermatan sulfate in cerebrospinal fluid as a biomarker for mucopolysaccharidosis II²
 - Rapid digestion, filtration, solid-phase extraction and chemical derivatization
- Case study 3: Development and validation of an LC-MS/MS method for the quantification of fascin proteins in human serum³
 - Immunocapture, protein precipitation, enzymatic digestion and solid phase extraction

^{2.} Peng Pan, Mu Chen, Zhiling Zhang, Amauri Dalla Corte, Carolina Souza, Roberto Giugliani, Luying Pan, Yongchang Qiu, Lakshmi Amaravadi, Jiang Wu. (2018) Bioanalysis

^{3.} Ke Li, Zhiling Zhang, Kai Wang, Xin-Yun Huang, Zhongping (John) Lin. (2022) Bioanalysis

Using LC-MS for biomarker analysis

When? How? Why?

Multiplexing

- Case study 4: Development and Validation of an LC-MS/MS Method for the Simultaneous Quantitation of Fifteen Bile Acids in Human Serum⁴
 - Fifteen bile acids, including five major bile acids (cholic acid, deoxycholic acid, ursodeoxycholic acid, chenodeoxycholic acid and lithocholic acid), and their respective glyco- and tauro- conjugates were extracted by protein precipitation using methanol.

Analyte	Statistics	QC LLOQ	Intra-run QC Low	Intra-run QC Mid	Intra-run QC High	Analyte	Statistics	QC LLOQ	Intra-run QC Low	Intra-run QC Mid	Intra-run QC High	Analyte	Statistics	QC LLOQ	Intra-run QC Low	Intra-run QC Mid	Intra-run QC High
CDCA	Inter-run %CV	10	10	3.7	3.5	GDCA	Inter-run %CV	6	6.4	5.1	6.1		Inter-run %CV	7.7	6.1	4.8	8.3
	Inter-run %Bias	8	0	-3.1	-5		Inter-run %Bias	2	-5.9	-4.4	-6.4	TCA	Inter-run %Bias	0	-6.7	-4.5	-1.7
CA	Inter-run %CV	9.2	6.2	5.2	4.6	GLCA	Inter-run %CV	17.1	9.6	4.1	3.3	TDCA	Inter-run %CV	14.8	9.6	4.7	4.6
	Inter-run %Bias	2	-3.3	-3.8	-3.1		Inter-run %Bias	-3.5	-5.7	-3.8	-6.3		Inter-run %Bias	-2.8	-4.7	-3.6	-6
DCA	Inter-run %CV	9	6.1	4.1	4.8	GUDCA	Inter-run %CV	11	6.7	4.2	3.7		Inter-run %CV	10	5.7	4.6	3.2
	Inter-run %Bias	-1.7	-1.3	-2.2	-4.2		Inter-run %Bias	-4	-4.3	-1.9	-6.8	TLCA	Inter-run %Bias	-3.2	-3	-5.8	-7.2
GCDCA	Inter-run %CV	6.6	4.7	5.2	2.9	LCA	Inter-run %CV	8	4.8	3.3	3.8	TUDCA	Inter-run %CV	14.6	7.1	5.1	5.9
	Inter-run %Bias	-3.5	1.3	-2.4	-8.3		Inter-run %Bias	1	-1	-1	-3.2		Inter-run %Bias	-6	-3.7	-3.9	-4
GCA	Inter-run %CV	4.8	6.1	4.2	4.1	TCDCA	Inter-run %CV	7.8	6.6	3.6	6.2	Int UDCA	Inter-run %CV	6.1	5.3	5	4.6
	Inter-run %Bias	-1.5	-2.3	-1	-6.5		Inter-run %Bias	3.5	-3.7	-5.1	-4.9		Inter-run %Bias	1	1.8	4	-2.3

4. Kai Wang, Siliang Chen, Yafei Xu, Luca Matassa, Zhongping (John) Lin, Pamela Vig, Hassan Rashidzadeh, Hongmei (Karen) Cao and Marita Larsson Cohen (2019) The American Association of Pharmaceutical Scientists (AAPS)







Aiying Yu, Ph.D.

Senior Scientist in Department of BioAnalytical Sciences (BAS) at Genentech since 2022. Ph.D. in Analytical Chemistry from Texas Tech University in 2022. Expertise in comprehensive structural identification and quantification of proteins, glycans, and glycopeptides in various biological matrices. Hands-on experience in analyzing tissues from Alzheimer's, breast cancer, and kidney brush-border membranes. Current work is focused on developing assays to enable characterization and quantification of protein therapeutics and biomarkers in fixed tissue samples such as formalin-fixed paraffin-embedded (FFPE) tissues using innovative LC-MS based approaches.



Target Occupancy in Frozen and FFPE Tissues using Immunoaffinity Capture 2D-LC-MS/MS

Aiying Yu, Lingyao Meng, Jintang He, Surinder Kaur, <u>Keyang Xu</u>

71th ASMS Conference, RBIG Workshop

June 7th, 2023

KRAS G12C and GDC-6036 Inhibitor

- KRAS G12C, a KRAS mutation favors the activated state of KRAS which results in uncontrolled cell growth and tumor formation.
- GDC-6036, an investigational KRAS G12C inhibitor that acts by irreversibly binding to the switch II pocket of KRAS G12C, blocking GTP binding and activation.





Purkey, Hans. Cancer Research 82.12_Supplement (2022): ND11-ND11.

Sensitive and Hybrid Assay (Immunoaffinity Capture + 2D-LC-MS/MS)



KRAS G12C Engagement of Xenograft Core Needle Tumor Biopsies



- Sub fmol/µg level sensitivity was achieved with 5 µg of total protein: ~ 5 10% of the core needle biopsy
- Increase trend of KRAS G12C engagement observed with dosing escalation

Lingyao Meng, Emily W. Chan, Carl Ng, et al.. Analytical Chemistry 2022 94 (37), 12927-12933

KRAS G12C Engagement of Mouse Xenograft FFPE Tissue



Summary:

- Several approaches were compared for assessment of KRAS G12C engagement in FFPE tissues.
- Several extraction buffers were evaluated and SDS buffer achieved the highest protein recovery.
- Early xenograft data indicates that target engagement in FFPE tissues comparable to that in frozen tissues.

Future work:

- Optimization for protein extraction and automation of sample preparation ongoing.
- Quantification of KRAS G12C engagement in FFPE tissues from clinical samples.

Acknowledgement

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- Suk Hyung
- Arun Taylor
- Helen Davis
- Sylvia Wong
- Jason Lamar

KRAS G12C Team

- Zhen Shi
- Emily W. Chen
- Mark Merchant

Pathology Labs

- Hartmut Koeppen
- Jian Jiang









Yifan Shi, Ph.D.

Senior Scientist in the Bioanalytical Discovery & Development Sciences department of Janssen R&D at Spring House, PA for 8 years. Associate director at Alliance Pharma (2011 – 2015). Staff scientist in Covance & Tandem Labs (2009 – 2011). PhD in chemistry from University of Utah. BSc in material chemistry from Peking University. Experience in small and large molecule quantitation, target engagement, and protein dynamics using various LC-MS platforms. Responsible for supporting drug discovery and development, including biomarker studies for TK and PK.



Implementing antibody-free strategies for protein quantitation using LC-MS

Yifan Shi, Ph.D.

Janssen R&D Spring House, PA June 7th, 2023 Colonic mucosa highlighted by Picro-Mallory trichrome special stain, from an exploratory study for Pulmonary Arterial Hypertension (PAH)

> Credit: Vini Carreira, Pathology, Preclinical Sciences & Translational Safety

Janssen Research & Development, LLC ©2022 JRD, LLC

Workflow for Hybrid LC-MS Protein Quantitation



Magnetic beads based immunoaffinity LC-MS protein quantitation

Advantages

- High throughput
- Relatively high sensitivity and dynamic range
- Hybrid LC-MS assay requirement
 - Recombinant protein
 - Capturing antibody

Potential issues

- Recombinant protein unavailable (or small fragments)
- Unreliable antibody supply

Antibody-free Protein Extraction and Digestion



FASP (Filter-Aid Sample Preparation) workflow

FASP basic concepts and advantages

- Ultrafiltration device retaining proteins
- Compatible with strong detergent for tissue homogenization and extraction
- Antibody-free and multiplex



High protein coverage for proteomics with FASP

FASP method applications

- Effective extraction of cytosolic and membrane proteins
- Multiple enzyme digestion to increase
 protein sequence coverage
- Efficient digestion at low protein levels

Antibody-free Protein Biomarker Quantitation

1500

1000-

500-

0-

LOQ (ng/ml)



High pH fractionation

- Orthogonal separation to identify low abundance target proteins
- Offline or online automation

Kim et al., Anal. Methods, 11, 4693, 2019 Zhang et al., Anal. Chem., 90, 1870, 2018



Microflow HPLC and online trapping

- Improved assay sensitivity
- Robust system for BA sample analysis

Case 1: FASP-Facilitated Method Development

Challenges

- Transmembrane protein with ~1300 AA
 - Recombinant full or fragment protein production failed at two vendors & Janssen internally
 - LC-MS method cannot be fully optimized without recombinant protein
- Low abundance in liver (<10 ng/g)
 - Difficulty in screening immunocapture antibodies using liver samples
 - No suitable reagent identified after testing 10 different antibodies

FASP for MS optimization & antibody screening

- Full length protein expression in a cell line
 - Adenovirus infection verified with mRNA detection
- FASP used for cell samples for LC-MS optimization
 - Highly sensitive LC-MS method developed for screening of antibody in liver homogenate for endogenous protein



LC-MS method optimization with AAV overexpressed cell samples processed using FASP

Case 2: FASP for Target Protein Bioanalysis

Background

Hybrid LC-MS method established

- Endogenous protein conc. ${\sim}11~\mu\text{g/g}$ in liver
- Study samples analyzed successfully
- Antibody discontinued by vendor
- Other antibodies have poor recovery or linearity problems for tissue samples

FASP for protein bioanalysis in liver homogenate

- Antibody-free target protein quantitation
- High correlation between immunocapture & FASP
- Excellent reproducibility
- Adequate sensitivity
- Lower throughput





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 - Brian Rady







Yang (Thomas) Tang, Ph.D.

Principal Scientist in Regulated Small Molecule Bioanalysis, DMPK at Genentech since 2020. Ph.D. in Analytical Chemistry from Boston University (2015-2020). At Genentech, responsible for developing and validating LC-MS/MS assays for quantitation of small molecular drugs and biomarkers to support TK and PK studies in pharmaceutical industry. Additional expertise in detailed structure characterization of metabolites and glycans using radical-induced MS/MS fragmentation methods.



A Universal Surrogate Matrix Assay for Urea Measurement in Clinical Pharmacokinetic Studies of Respiratory Diseases

Yang (Thomas) Tang Jun. 7th, 2023

Urea Bioanalysis



- Physiological matrices in respiratory diseases: Nasal secretion (NS), exhaled breath condensate (EBC), mucosal lining fluid (MLF), lung epithelial lining fluid (ELF) and plasma/serum.
- The measurement of drug concentration is challenging (Volume unknown).
- Urea as <u>Dilution Marker</u> to normalize liquid sample volumes [Urea] is consistent in most matrices from the same individual.
- 1. Diffuse freely and quickly throughout the body.
- 2. Low turnover rate.
- 3. Minimally affected by disease states.

Urea quantitation by LC-MS/MS

- High specificity, sensitivity and easy accessibility.
- Inherent difficulties:
 - a) Vulnerable to interference.
 - b) High background.
 - c) Endogenous presence.



 $V_{native} = (C_{buffer eluate}/C_{plasma}) \times V_{buffer eluate}$

Approaches:

- 1) Chemical derivatization.
- 2) Surrogate analyte.
- 3) Surrogate matrix.

Surrogate Analyte vs Surrogate Matrix





Surrogate Analyte

- Same biological matrix.
- [¹⁵N₂]-urea as calibration curve.
- Signal ratio of urea and [¹⁵N₂]-urea predetermined.

Assay issues:



One assay per matrix

- Cumbersome data processing.
- Varied signal ratio.
- Endogenous QCs fail.



Improvable

Assay Issues:

A universal assay for

- Poor LLOQ peak shape. all matrices
- High baseline.

Comparison of Ionization Modes and Platforms



Sciex 6500

Sciex 4500

Sciex 5500



Neat solution injected



Surrogate Matrix Suitability



Surrogate matrix:

- 1) Contain no measurable endogenous analyte.
- 2) Lack matrix effects or interferences.

Parallelism

Parallelism of calibration curves between native and surrogate matrices.

Endogenous QCs

Quantitation accuracy of EQCs by surrogate matrix curve.



Minimal matrix effects have been confirmed by good parallelism between curves prepared in water, saline, diluted plasma and diluted serum.

Additional Tests



Accuracy and Precision

Sample	Theoretical (μg/mL)	Calculated (µg/mL)	RSD (%)	Accuracy (%)
Cal 1	1	1.01	10.2	101.3
Cal 2	2	1.94	2.0	96.8
Cal 3	4	4.05	4.9	101.2
Cal 4	10	10.04	4.1	100.4
Cal 5	15	14.54	1.4	96.9
Cal 6	25	26.06	2.8	104.2
Cal 7	45	45.02	2.9	100.0
Cal 8	50	49.57	1.5	99.1
LQC	3	2.91	3.0	97.1
MQC	12	12.03	2.2	100.2
HQC	40	39.97	3.7	99.9
DQC (20×)	300	307.01	3.1	102.3
Plasma ELQC (5×)	50	50.45	1.8	100.9
Plasma EQC (20×)	322	329.77	2.8	102.4
Plasma EHQC (50×)	1000	1045.59	1.6	104.6
Serum ELQC (5×)	50	48.39	1.8	96.8
Serum EQC (20×)	274	280.27	3.7	102.3
Serum EHQC (50×)	1000	1020.52	2.4	102.1

Surrogate Analyte vs Surrogate Matrix

\neg
c y (%)
).9
.4
.6
.8
3
2.1
0 2 4 2 2

Comparable results demonstrate the satisfying performance of the newly developed surrogate matrix assay.

- Recovery
- Sensitivity
- Benchtop Stability (RT, 24 hr)

EQC: Endogenous QC ELQC: Endogenous low QC EHQC: Endogenous high QC

The newly developed assay is robust and reliable.

Conclusion



- A reliable LC-MS/MS surrogate matrix assay has been developed and qualified for urea measurement in plasma, serum, or BALF (bronchoalveolar lavage fluid).
- Sciex Triple Quad 5500 with ESI was identified as the most suitable platform.
- LC: isocratic elution with salt-free MP to give robustness and tolerability of interferences.
- MS: parameters were comprehensively optimized to maximize S/N.
- Saline was identified as the surrogate matrix. Good parallelism indicates minimal matrix effects.
- The assay demonstrated good precision and accuracy, dilution integrity (50×), sensitivity, recovery, and stability.
- The new surrogate matrix assay is easier to operate than the surrogate analyte assays, and more specific and accurate than colorimetric assays.

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THANK YOU



Questions (1)



- For acceptance criteria of Multiplex Biomarker analysis, would keep the same as singelicate or different?
- For data analysis of Multiplex Biomarker, any existing or new tool that can be used to make the work more efficient?



Questions (2)



- What are the advantages to using the SDS buffer over MS-friendly buffers such as RapiGest for protein extraction in tissue samples?
- How to determine the retention time window for targeted peptides in a 2D-LC system?



Questions (3)



- What types of biological matrices have you used this approach for bioanalytical studies? How difficult is it to develop methods in different matrices for the same protein target?
- What are the considerations for selecting an internal standard for FASP methods?





- What are the advantages and disadvantages of using surrogate analyte vs surrogate matrix in LC-MS biomarker analysis?
- In your organization, how many biomarker assays are qualified and how many are validated? What's the considerations in determining qualified or validated?
- In surrogate matrix assay development, when evaluating the parallelism between the surrogate matrix and native matrix, what is the typical acceptance criteria?
- Instrument platforms, Triple quadrupole vs. HR-MS?



Workshop Summary



- The workshop with ~70 attendees was featured by four excellent short presentations by the experts in the field of biomarker analysis using mass spectrometry, which was followed by panel discussions and Q&A on wide-range of important topics.
- The audience appreciated the topics, insights, opinions, and experiences shared by the panels and attendees, including experts (e.g., Hendrik Neubert, Dawn Dufield) in the field.
- Positive feedbacks were received from the audience on the quality and coverage of the presentations and discussions.